AMPure Bead Cleanup

Required equipment:
Vortex mixer with a foam tube holder attachment (similar to VWR Part#12620-876), or a rotator.
Magnetic tube rack
Ampure XP beads
Lo-Bind tubes
Freshly made 70% ethanol

- *Tips: You will get a better yield if you increase the volume of ethanol for the washes enough to fill the tube. To recover lost DNA from supernatant, do a 1X bead wash.*
- Let the AMPure beads equilibrate to room temp and vortex to resuspend completely.
- To increase yield, use Lo-Bind tubes for your sample(s).

For:	**100bp**	**300-500bp**
Add:	1.8X volume of beads	1.0X volume of beads

- Mix the DNA and beads thoroughly by pipetting or inverting the tube.
- Pulse-spin the tube briefly. Do not pellet the beads.
- Bind the DNA to the beads by shaking on the vortex mixer, using the blue foam top, at setting 7 (2000 rpm) for 10 minutes (or place on a rotator).
- Pulse-spin the tube briefly. Do not pellet the beads.
- Place the tube on the magnetic rack for at least 3 minutes, until the supernatant clears.
- With the tube still on the rack, remove and save the supernatant (in case of error). It is better to leave buffer behind than to lose some of your beads.
- With the tube still on the rack, add 200µl -1 mL of freshly-prepared 70% ethanol, enough to cover the beads.
- Wait 30 seconds, then remove and discard the ethanol.
- Repeat the last two steps once.
- Pulse-spin the tube to collect any remaining ethanol at the bottom. Put it back on the magnetic rack and pipette off the remaining ethanol.
- Allow tube to air-dry for up to 1 minute.
- Add 20-35µl of EB or ultrapure water and pipette up and down to resuspend.
- Place on the vortex shaker for 2-10 minutes (or on the rotator for 10 minutes).
- Pulse-spin the tube and place it back on the magnetic rack.
- After ~30 seconds, when the supernatant is clear, transfer the eluted DNA to a clean Lo-Bind tube.